

Oligonucleotide Properties Calculator

Enter Oligonucleotide Sequence Below	
OD and Molecular Weight calculations are for single-stranded DNA	
Nucleotide base codes ATT GAA ACA CAA AAT ACC AGT TCT CAA ATA CAA TGA ACA TTA TTA ATT ATA ATT CAG TTA AAA GTC ATT GAT CAG AAC AGC AGT GAA GGT TAG CTA TAA GCG CGT TAT AGG TGC AGG CAG AGT GTC GTG CCT ATA TAT ACC CTT TGS AAT GCA CAA GTT GAA CAC AAA GAA AAA TG	
Reverse Complement Strand(5' to 3') is: CAT TTT TCT TTG TGT TCA ACT TGT GCA TTC CAA AGG GTA TAT ATA GGC ACG ACA CTC TGC CTG CAC CTA TAA CGC GCT TAT AGC TAA CCT TCA CTG CTG TTC TGA TCA ATG ACT TTT AAC TGA ATT ATA ATT AAT AAT CTT CAT TGT ATT TGA GAA CTG GTA TTT TGT GTT TCA AT	
Number of Fluorescent tags per strand: 0 6-FAM 0 TET 0 HEX 0 TAMRA	
Minimum base pairs required for single primer self-dimerization: 5	
Minimum base pairs required for a hairpin: 4	
<input type="button" value="Calculate"/> <input type="button" value="SWAP STRANDS"/> <input type="button" value="BLAST2"/> <input type="button" value="Check Self-Complementarity"/>	
Physical Constants	
Length: 176 bases GC content: 35 % Molecular Weight: 54503.7 1 ml of a sol'n with an Absorbance of 1 at 260 nm is .493 microMolar and contains 26.9 micrograms	
Melting Temperature (T _M) Calculations	
1 75 °C (Basic) 2 89 °C (Salt Adjusted) 3 79 °C (Nearest Neighbor) 50 nM Primer 50 mM Salt (Na ⁺)	
Thermodynamic Constants	
Conditions: 1 M NaCl at 25°C at pH 7.	
RlogK 33.404 cal/(°K*mol) deltaH 1409.4 Kcal/mol deltaG 246.1 Kcal/mol deltaS 3735.1 cal/(°K*mol)	

To use this calculator, you must be using Netscape 3.0 or later or Internet Explorer version 3.0 or later, or another Javascript-capable browser

Self-Complementarity requires a 4.x browser. IE 5.0 is also supported.

This page was written in Javascript.

Extensively rewritten from 12/15/2000-12/19/2000 to isolate javascript

Oligo object behaviors for teaching purposes.

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About the Calculations

Thermodynamic Calculations

The nearest neighbor and thermodynamic calculations are done essentially as described by Breslauer *et al.*, *Proc. Nat. Acad. Sci.* **83**, 3746-50, 1986 ([Abstract](#)) but using the values published by Sugimoto *et al.*, *Nucl. Acids Res.* **24**, 4501-4505, 1996 ([Abstract](#)). This program assumes that the sequences are not symmetric and contain at least one G or C. The minimum length for the query sequence is 8.

The melting temperature calculations are based on the simple thermodynamic relationship between entropy, enthalpy, free energy and temperature, where

$$\Delta H = \Delta G + T\Delta S$$

The change in entropy (order or a measure of the randomness of the oligonucleotide) and enthalpy (heat released or absorbed by the oligonucleotide) are directly calculated by summing the values for nucleotide pairs obtained by Breslauer *et al.*, *Proc. Nat. Acad. Sci.* **83**, 3746-50, 1986. The relationship between the free energy and the concentration of reactants and products at equilibrium is given by

$$\Delta G = RT \ln \left(\frac{[DNA \cdot primer]}{[DNA][primer]} \right)$$

Substituting the two equations gives us

$$\Delta H = T\Delta S + RT \ln \left(\frac{[DNA \cdot primer]}{[DNA][primer]} \right)$$

and solving for temperature T gives

$$T = \frac{\Delta H}{\Delta S + R \ln \left(\frac{[DNA \cdot primer]}{[DNA][primer]} \right)}$$

We can assume that the concentration of DNA and the concentration of the DNA-primer complex are equal, so this simplifies the equation considerably. It has been determined empirically that there is a 5 (3.4 by Sugimoto et al.) kcal free energy change during the transition from single stranded to B-form DNA. This is presumably a helix initiation energy. Finally, adding an adjustment for salt gives the equation that the Oligo Calculator uses:

$$T = \frac{\Delta H - 5 \frac{\text{kcal}}{^\circ\text{K mole}}}{\Delta S + R \ln \left(\frac{1}{[\text{primer}]} \right)} + 16.6 \log_{10} ([\text{Na}^+])$$

No adjustment constant for salt concentration is needed, since the various parameters were determined at 1 Molar NaCl, and the log of 1 is zero.

ASSUMPTIONS:

The thermodynamic calculations assume that the annealing occurs at pH 7.0. The melting temperature (T_m) calculations assume the sequences are not symmetric and contain at least one G or C. The oligonucleotide sequence should be at least 8 bases long to give reasonable T_m s.

Basic Melting Temperature (T_m) Calculations

The two standard approximation calculations are used. For sequences less than 14 nucleotides the formula is

$$T_m = (wA + xT) * 2 + (yG + zC) * 4$$

where w,x,y,z are the number of the bases A,T,G,C in the sequence, respectively.

For sequences longer than 13 nucleotides, the equation used is

$$T_m = 64.9 + 41 * (yG + zC - 16.4) / (wA + xT + yG + zC)$$

ASSUMPTIONS:

Both equations assume that the annealing occurs under the standard conditions of 50 nM primer, 50 mM Na^+ , and pH 7.0.

Salt Adjusted Melting Temperature (Tm) Calculations

A variation on two standard approximation calculations are used. For sequences less than 14 nucleotides the same formula as the basic calculation is used, with a salt concentration adjustment

$$T_m = (wA + xT) * 2 + (yG + zC) * 4 - 16.6 * \log_{10}(0.050) + 16.6 * \log_{10}([Na^+])$$

where w,x,y,z are the number of the bases A,T,G,C in the sequence, respectively.

The term $16.6 * \log_{10}([Na^+])$ adjusts the Tm for changes in the salt concentration, and the term $\log_{10}(0.050)$ adjusts for the salt adjustment at 50 mM Na⁺. Other monovalent and divalent salts will have an effect on the Tm of the oligonucleotide, but sodium ions are much more effective at forming salt bridges between DNA strands and therefore have the greatest effect in stabilizing double-stranded DNA.

For sequences longer than 13 nucleotides, the equation used is

$$T_m = 100.5 + (41 * (yG + zC) / (wA + xT + yG + zC)) - (820 / (wA + xT + yG + zC)) + 16.6 * \log_{10}([Na^+])$$

Symbols and salt adjustment term as above, with the term $(41 * (yG + zC - 16.4) / (wA + xT + yG + zC))$ adjusting for G/C content and the term $(820 / (wA + xT + yG + zC))$ adjusting for the length of the sequence.

ASSUMPTIONS:

Both equations assume that the annealing occurs under the standard conditions of 50 nM primer and pH 7.0.

OD Calculations

Molar Absorptivity values in 1/(Moles cm)

Residue	Moles ⁻¹ cm ⁻¹	Molecular Weight (after protecting groups are removed)
Adenine (dAMP, Na salt)	15200	313.21
Guanine (dGMP, Na salt)	12010	329.21
Cytosine (dCMP, Na salt)	7050	289.18
Thymidine (dTMP, Na salt)	8400	304.2
6-FAM	20960	537.46
TET	16255	675.24
HEX	31580	744.13
TAMRA	31980	

Assume 1 OD of a standard 1ml solution, measured in a cuvette with a 1 cm pathlength.

6-FAM:

Chemical name:	6-carboxyfluorescein
Absorption wavelength maximum:	495 nm
Emission wavelength maximum:	521 nm
Molar Absorptivity at 260nm:	20960 Moles ⁻¹ cm ⁻¹

TET:

Chemical name:	4, 7, 2', 7'-Tetrachloro-6-carboxyfluorescein
Absorption wavelength maximum:	519 nm
Emission wavelength maximum:	539 nm
Molar Absorptivity at 260nm:	16255 Moles ⁻¹ cm ⁻¹

HEX:

Chemical name:	4, 7, 2', 4', 5', 7'-Hexachloro-6-carboxyfluorescein
Absorption wavelength maximum:	537 nm
Emission wavelength maximum:	556 nm
Molar Absorptivity at 260nm:	31580 Moles ⁻¹ cm ⁻¹

TAMRA:

Chemical name:	N, N, N', N'-tetramethyl-6-carboxyrhodamine
Absorption wavelength maximum:	555 nm
Emission wavelength maximum:	580 nm
Molar Absorptivity at 260nm:	31980 Moles ⁻¹ cm ⁻¹

Nucleotide base codes (IUPAC)

Symbol: nucleotide(s)		
A adenine	M A or C	K G or t
C cytosine	R A or G	V A or C or G; not T
G guanine	W A or T	H A or C or T; not G
T thymine in DNA; uracil in RNA	S C or G	D A or G or T; not C
N A or C or G or T	Y C or T	B C or G or T; not A

Most recent version is available at URL: <http://www.basic.northwestern.edu/biotools/oligocalc.html>

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